

Water-Soluble Polypeptides with Elongated, Charged Side Chains Adopt Ultrastable Helical Conformations

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The α -helix is one of the most important structural domains in polypeptides controlling numerous biological activities and functions.^{1–12} Studies aimed at increasing the overall helicity and stability of helical motifs of peptides, especially oligopeptides, have contributed to the fundamental understanding of peptide folding/unfolding and have led to improvements of their biological and pharmaceutical activities.^{13–19} There is often a drawback in the design of water-soluble, bioactive helical peptides: charged amino acid building blocks provide water solubility but decrease helicity because of disruption of helix due to side-chain charge repulsion.^{20–23} Increasing the proportion of hydrophobic amino acids tends to increase helicity by increasing side-chain hydrophobic interactions, but the resulting structures show reduced water solubility, which is undesirable for the design of biologically active peptides. It has been a general strategy to integrate both water-soluble and helix-stabilizing motifs in the peptide structure to design water-soluble, helical peptides. Such peptides are often designed to have charged amino acid residues situated on one side of the helix surface and the residues responsible for stabilizing the helix through side-chain hydrophobic interactions,^{24–26} salt bridges,^{27–29} or tethering^{18,30,31} situated on the opposite side of the helix surface. These strategies require the design of peptides with specific sequences²⁷ and usually involve tedious chemistries of polypeptide side chains³⁰ that are typically difficult to control. For polypeptides prepared by polymerization instead of through stepwise synthesis, such helix-stabilization strategies mentioned above for the synthesis water-soluble, helical peptides cannot be simply applied.³²

Water-soluble, synthetic polypeptides that can adopt stable α -helical conformations have attracted much attention. Prior efforts have been focused on introducing neutrally charged, hydrophilic functional groups³³ or moieties.³² Poly(*N*-hydroxyalkyl-L-glutamine),³³ one of the early design of water-soluble polypeptides derived from aminolysis of poly(L-glutamate) (PLG) with pendant hydroxyl groups, showed excellent water-solubility and fairly high helical contents (up to ca. 65% helicity) in aqueous solution.³³ Later, Deming and co-workers designed poly(L-lysine) (PLL) containing pendant oligoethylene glycol moieties.³² The resulting oligoethylene glycol-graft PLL showed excellent water solubility and remarkably high helical content (100% helicity in pH 7 water at 25 °C). Recently, Li and co-workers designed thermo-responsive α -helical polypeptides from peglated PLG, highlighting the recent progress of this class of special polypeptides

containing noncharged, water-soluble segments on a α -helical structures.³⁴

In a separate effort, we designed charged, water-soluble polypeptides that adopt stable α -helical conformations (i.e., α -helical polypeptide electrolytes; α HPEs), by using polypeptide containing charged side chains but elongating the charge-containing amino acid side chains to place the charges distally from the polypeptide backbone (Figure 1a).³⁵ When the charges are 11 σ -bonds away from the peptide backbone, as in poly(γ -(4-(1-hexanol-6-aminomethyl))benzyl-L-glutamate) (PVBLG-1; Figure 1b), the resulting polypeptide with a degree of polymerization (DP) of 60 ((PVBLG-1)₆₀) maintains a stable α -helical conformation with 91% helicity.³⁵ PVBLG-1's with very low DPs, such as (PVBLG-1)₁₀ with a DP value of 10, however, have mixed conformations containing both β -sheets and α -helices, with a helicity of only 26% for (PVBLG-1)₁₀.³⁵ Because both the charge-backbone distance and the hydrophobicity of the side chains in α HPEs have significant effect on the stability of α -helix,³⁵ we reasoned that further elongating the side chain will not only further reduce side chain charge repulsion by increasing the charge-backbone distance but also increase the side-chain hydrophobicity. By doing so, it is possible to obtain a water-soluble α HPE with ultrastable α -helix and high helicity even at a very low DP. Here, we report the design and synthesis of a water-soluble α HPE, (poly(γ -(4-aminoethylthiopropoxy)benzyl-L-glutamate) (PAOBLG-AET, Figure 1c), with side chain charge situated 17 σ -bonds away from the peptide backbone, adopts an unprecedented, remarkably high helicity (81%) with a DP of 10 at pH 2 aqueous solution.

The synthesis of PAOBLG-AET is illustrated in Figure 1d. γ -(4-Allyloxylbenzyl)-L-glutamate *N*-carboxyanhydride (AOB-L-Glu-NCA) can be easily prepared in multigram scale (see Supporting Information). The ring-opening polymerizations (ROPs) of AOB-L-Glu-NCA initiated by hexamethyldisilazane (HMDS)^{36–38} yielded PAOBLGs with controlled molecular weights (MWs) and narrow molecular-weight distributions (MWDs) that were determined by gel permeation chromatography (GPC) (Table S1, Supporting Information). For example, at a monomer/initiator (M/I) ratio of 10 with expected M_n of $3.0 \times 10^3 \text{ g} \cdot \text{mol}^{-1}$, the resulting PAOBLG had an M_n of $2.8 \times 10^3 \text{ g} \cdot \text{mol}^{-1}$ with a

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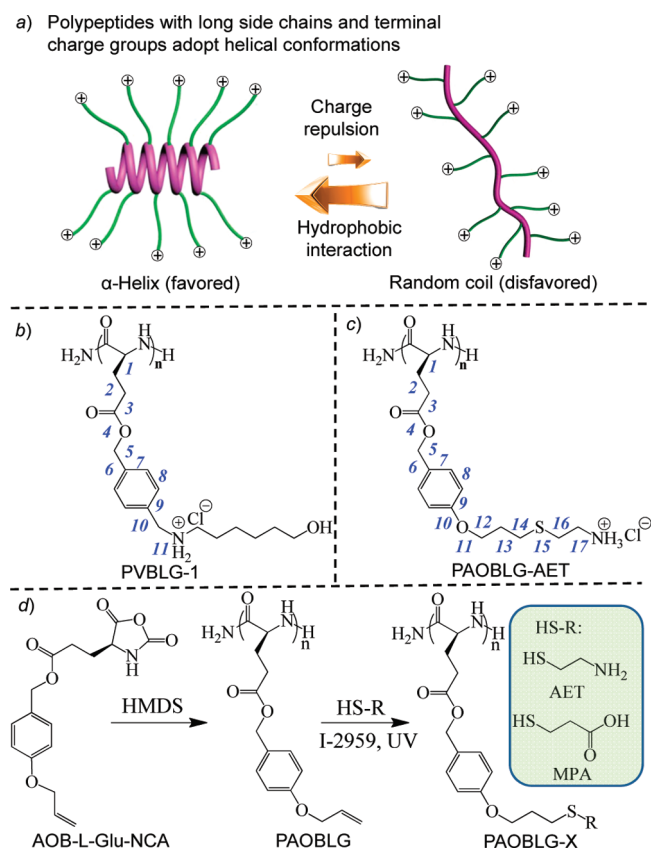


Figure 1. (a) Polypeptide with charged side chains and the postulated helix–coil transition in response to the length of the side chains. Chemical structures of (b) PVBLG-1 and (c) PAOBLG-AET. (d) Synthesis of PAOBLG-AET and PAOBLG-MPA.

narrow MWD of 1.22 (entry 1, Table S1, Supporting Information). The MW and MWD of PAOBLG₁₀ obtained by matrix-assisted laser desorption ionization mass spectrometry (MALDI–TOF MS, Figure S3, Supporting Information) agreed well with the values obtained by GPC. To accelerate the polymerization of AOB-L-Glu-NCA and synthesize higher MW PAOBLGs, we used 1,5,7-triazabicyclo[4.4.0]dec-5-ene (TBD) as a cocatalyst,^{39,40} which gave faster yet controlled NCA polymerization (Figure S4, Supporting Information).⁴¹ In the presence of a small amount of TBD (HMDS/TBD = 1/0.1), the polymerizations yielded corresponding PAOBLGs with the expected MWs and narrow MWDs (entries 3 and 4, Table S1, Supporting Information).

The PAOBLGs were then treated with 2-aminoethanethiol hydrochloride in a mixture of dimethylformamide and deionized water to effect a UV-triggered thiol–ene “click” reaction.^{42–46} Dialysis of the reaction mixture followed by lyophilization removed all the small-molecule impurities and afforded the desired polymers as a fluffy powder. As expected, the thiol–ene reaction proceeded rapidly and completed in 10 min, yielding PAOBLG-AETs with nearly quantitative grafting efficiency (Figures S5 and S6, Supporting Information).

The PAOBLG-AETs are very soluble in water (>20 mg·mL⁻¹) because of the terminal ammonium groups on each of their side chains, in sharp contrast to PAOBLG which is insoluble in water. To determine whether the PAOBLG-AETs have the expected high helicity at low DP, we used circular dichroism (CD) spectroscopy

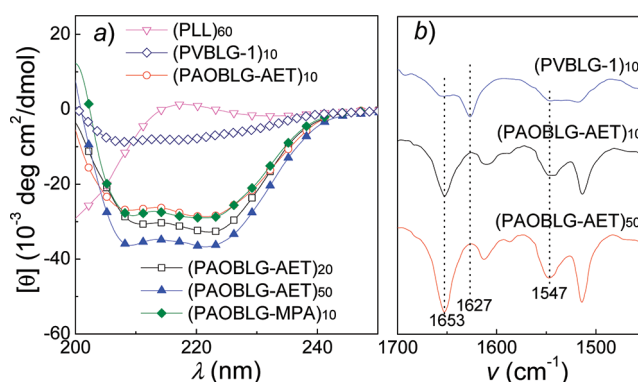


Figure 2. (a) CD spectra of various polypeptides bearing charged side chains ((PLL)₆₀, (PVBLG-1)₁₀ and (PAOBLG-AET)_{10,20,50}) in aqueous solution at pH 2 and (PAOBLG-MPA)₁₀ in aqueous solution at pH 10. (b) Fourier-transform infrared spectra (FTIR) of (PVBLG-1)₁₀ and (PAOBLG-AET)₁₀ and (PAOBLG-AET)₅₀.

to analyze the conformation of the PAOBLG-AETs at pH 2 at which all side-chain amines should be protonated and are charged. All three PAOBLG-AETs ((PAOBLG-AET)₁₀, (PAOBLG-AET)₂₀, and (PAOBLG-AET)₅₀) showed the characteristic CD spectra of α -helix with two minima at 208 and 222 nm (Figure 2a). (PAOBLG-AET)₁₀ (charge–backbone distance of 17 σ -bonds, Figure 1c) had a $-\theta_{222}$ value of $28.5 \times 10^3 \text{ cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$, which corresponds to a helicity of 81% (Figure 2a, Table 1),⁴⁷ in sharp contrast to a 60-mer poly(L-lysine) ((PLL)₆₀, charge–backbone distance of 4 σ -bonds) that adopts a random coil conformation and (PVBLG-1)₁₀ (charge–backbone distance of 11 σ -bonds, Figure 1b) that has a $-\theta_{222}$ value of $7.2 \times 10^3 \text{ cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$, which corresponds to a helicity of only 26% (Figure 1a and Table 1). The high helicity of (PAOBLG-AET)₁₀ was further verified by FTIR (Figure 2b).^{48,49} (PVBLG-1)₁₀ has mixed conformations containing both α -helix (amide I band at 1653 and amide II band 1547 cm^{-1}) and β -sheet (amide I band at 1627 cm^{-1}) in solid state, while (PAOBLG-AET)₁₀ has predominant α -helix (strong amide I band at 1653 and amide II band 1547 cm^{-1}) and negligible β -sheet conformation. For PAOBLG-AETs with DP values of 20 and 50, the $-\theta_{222}$ values were 34.0 and $36.8 \times 10^3 \text{ cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$, corresponding to helicities of 94% and 100%, respectively (Figure 2a, and Table 1). (PAOBLG-AET)₁₀ and (PAOBLG-AET)₅₀ have nearly identical FTIR spectra (Figure 2b), further validating the high helical content of (PAOBLG-AET)₁₀.

We next studied the helical stability of PAOBLG-AETs against variation of environmental conditions, including changes in the pH and temperature and the presence of denaturing reagents. The $-\theta_{222}$ value of (PAOBLG-AET)₁₀ remained unchanged when the solution's pH was increased from 1 to 8 (Figure 3a). At further increased pH values, (PAOBLG-AET)₁₀ became less soluble because of deprotonation of some of its charged ammonium groups. (PAOBLG-AET)₁₀ showed a lack of concentration dependence of its $-\theta_{222}$ value in helix-forming conditions, suggesting that it remained monomeric in aqueous solution (Figure S8, Supporting Information). It displayed excellent helical stability against elevated temperature, with its $-\theta_{222}$ value decreasing 25% from $28.8 \times 10^3 \text{ cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$ at 4 $^\circ\text{C}$ to $21.6 \times 10^3 \text{ cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$ at 70 $^\circ\text{C}$ (Figure 3b), and against helix-destabilizing conditions such as high concentrations of NaCl (Figure 3c) and urea (Figure 3d). (PAOBLG-AET)₁₀ showed

Table 1. Conformation Analysis of Ionic Polypeptides

entry	polypeptide	DP	$-\langle[\theta]_{222} \times 10^{-3}\rangle$ ($\text{cm}^2 \cdot \text{deg} \cdot \text{dmol}^{-1}$) ^a	helical content (%) ^b
1	(PLL) ₆₀	60	-	0
2	(PVBLG-1) ₁₀	10	7.2	26
3	(PAOBLG-AET) ₁₀	10	28.5	81
4	(PAOBLG-AET) ₂₀	20	34.0	94
5	(PAOBLG-AET) ₅₀	50	36.8	100
6	(PAOBLG-MPA) ₁₀	10	29.6	84

^aThe mean residue molar ellipticity $\langle[\theta]\rangle$ was determined by following literature-reported formula: Ellipticity ($\langle[\theta]_{222}$ in $\text{cm}^2 \text{ deg dmol}^{-1}$) = (millidegrees \times mean residue weight)/(path length in millimeters \times concentration of polypeptide in $\text{mg} \cdot \text{mL}^{-1}$). ^bThe helical contents of the polypeptides were calculated using the following equation: percentage of α -helix = $(-\langle[\theta]_{222} + 3000)/39\,000$.⁴⁷

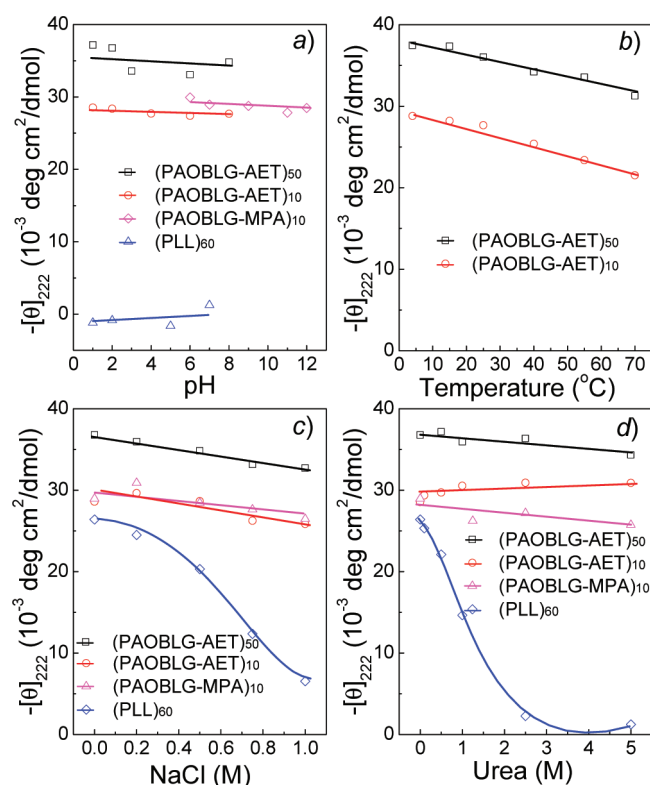


Figure 3. (a) pH dependence of residue molar ellipticity at 222 nm for (PAOBLG-AET)₁₀, (PAOBLG-AET)₅₀, (PAOBLG-MPA)₁₀, and (PLL)₆₀ at 0.05 $\text{mg} \cdot \text{mL}^{-1}$. (b) Temperature dependence of residue molar ellipticity at 222 nm for (PAOBLG-AET)₁₀ and (PAOBLG-AET)₅₀ at pH 2 and 0.05 $\text{mg} \cdot \text{mL}^{-1}$. (c) Salt-concentration dependence of residue ellipticity at 222 nm for (PAOBLG-AET)₁₀ and (PAOBLG-AET)₅₀ at pH 2 and (PAOBLG-MPA)₁₀ and (PLL)₆₀ at pH 10 ($c = 0.05 \text{ mg/mL}$). (d) Helical stabilities of (PAOBLG-AET)₁₀ and (PAOBLG-AET)₅₀ at pH 2, and (PAOBLG-MPA)₁₀ and (PLL)₆₀ at pH 10 in the presence of urea ($c = 0.05 \text{ mg/mL}$).

unprecedented helical stability against any known α -peptides and amazingly maintained $\sim 100\%$ of its original helical content in 5 M urea. (PAOBLG-AET)₅₀ showed very similar helical stability as (PAOBLG-AET)₁₀ to those changing environmental conditions; the helical stabilities of both (PAOBLG-AET)₁₀ and

(PAOBLG-AET)₅₀ were drastically different from that of PLL₆₀ in high concentrations of NaCl (Figure 3c) and urea solutions (Figure 3d).

This novel strategy of distal charge placement on side chains to maintain both water solubility and high helicity in low MW polypeptide can also be extended to polypeptides bearing negatively charged side chains. (PAOBLG-MPA)₁₀, a peptide with similar structure as (PAOBLG-AET)₁₀ bearing carboxylate terminated side-chain with charge-backbone distance of 18 σ -bonds, was prepared via thiol-ene reaction of PAOBLG with 3-mercaptopropionic acid (Figures 1d and S7, Supporting Information). (PAOBLG-MPA)₁₀ had a helicity of 84% in aqueous solution at pH 9, when its carboxylate groups were completely deprotonated. The $-\langle[\theta]_{222}$ value of (PAOBLG-MPA)₁₀ remained unchanged when the solution's pH was decreased from 12 to 6 (Figure 3a). At further decreased pH values, (PAOBLG-MPA)₁₀ became less soluble because of protonation of some of its charged carboxylate ions. (PAOBLG-MPA)₁₀ showed very similar response as (PAOBLG-AET)₁₀ against the helix-destabilizing conditions such as high concentrations of NaCl (Figure 3c) and urea (Figure 3d).

In summary, polypeptides with long side chain bearing charge groups were synthesized by controlled ROP of AOB-L-Glu-NCA and subsequent thiol-ene reactions. Because of their elongated hydrophobic side chains and distally situated charges, these polypeptides are highly water-soluble and have very high helicity even with a DP value as low as 10. Furthermore, the helical structures of these low MW polypeptide electrolytes were stable against changes in pH, temperature, and salt and urea concentrations. To our knowledge, PAOBLG-AET(MPA) is the shortest, charged peptide to show such high helicity, remarkable helical stability and water solubility. Our study demonstrates that elongating the hydrophobic side chain bearing a terminal charge group can serve as a general strategy for the design of water-soluble polypeptide with high helicity and high helical stability.

ASSOCIATED CONTENT

Supporting Information. Experimental details and the spectroscopy and analytical data for the synthesis and characterization of PAOBLG-AET and PAOBLG-MPA. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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